

Article

Placental mesenchymal and cord blood stem cell therapy for dilated cardiomyopathy



Neil Riordan is President and CEO of Medistem Laboratories. He directed a leading cancer laboratory for 10 years, Project RECNAAC of The Centre for the Improvement of Human Functioning, and he has proved himself a successful entrepreneur. In 2003 Dr Riordan became the director of research at the Immunology Research Centre. His research findings have been published in many prestigious journals, including *British Journal of Cancer* and *Medical Hypotheses*. His educational qualifications include MUA, PhD, MS, and PA from University of Nebraska, College of Medicine, and BS *magna cum laude* from Wichita State University.

Dr Neil Riordan

Thomas E Ichim¹, Fabio Solano², Roberto Brenes², Eduardo Glenn², Junbiao Chang³, Kyle Chan¹, Neil H Riordan^{1,3,4}
¹Medistem Laboratories Inc., 2027 E Cedar Street Suite 102 Tempe, AZ 85281, USA; ²Institute for Cellular Medicine, San Jose, Costa Rica; ³Zhengzhou University, Zhengzhou, China
⁴Correspondence: Tel: +1 954 7273662; Fax: +1 954 2060637; e-mail: riordan@medisteminc.com

Abstract

Regenerative treatment of dilated, non-ischaemic cardiomyopathy represents a significant unmet clinical need. Intracoronary administration of autologous bone marrow stem cells has demonstrated positive results in treatment of post-infarct and chronic ischaemic patients. Limitations of this procedure include: invasiveness of bone marrow extraction and cardiac catheterization, and dependence on stem cell populations that are aged and possibly senescent. Here, the use of intravenously administered allogeneic placental matrix derived mesenchymal stem cells for treatment of dilated cardiomyopathy is discussed. Safety of this cell population has already been established in completed Phase I and II trials; however, to date, clinical implementation for dilated cardiomyopathy has not been reported. Preclinical studies have demonstrated that mesenchymal stem cells: (i) inhibit myocardial inflammation; (ii) inhibit cardiomyocyte apoptosis; (iii) stimulate angiogenesis; and (iv) display therapeutic activity in models of dilated cardiomyopathy. Clinical studies have demonstrated the ability of mesenchymal stem cells to inhibit post-infarct remodelling, as well as potentially block inflammatory processes in graft versus host and Crohn disease. Presented here is case report of a patient with dilated cardiomyopathy treated with intravenous allogeneic mesenchymal stem cells and expanded umbilical cord blood CD34 cells who underwent a profound clinical improvement.

Keywords: angiogenesis, CD34, cord blood, dilated cardiomyopathy, mesenchymal, stem cell therapy

Introduction

Stem cell therapy in the post-myocardial infarct setting has yielded positive results in preclinical (Orlic *et al.*, 2001; Kudo *et al.*, 2003) and clinical studies (Chen *et al.*, 2004; Fernandez-Aviles *et al.*, 2004; Osterziel, 2007). However, the promise of regenerating myocardium appeals to other non-acute cardiac degenerative conditions besides myocardial infarction. For example, dilated cardiomyopathy is believed to have an annual incidence of approximately 400,000 per year in the USA alone, with a 50% survival at 5 years. Current therapies for this condition such as angiotensin-converting enzyme inhibitors, beta-blockers and vasodilators seek to reduce the rate at which myocardial damage accrues but do not have regenerative potential. This article begins by reviewing the disease and pathological mechanisms associated with its progression, and subsequently reviews various types of stem cell therapies

that have demonstrated promise in cardiac degeneration in general and current work in dilated cardiomyopathy. Although embryonic stem cells have previously been demonstrated to possess cardiogenic potential (van de Stolpe *et al.*, 2005), due to both technical and ethical limitations (Beeson and Lippman, 2006; Findikli *et al.*, 2006), these cells will not be discussed in this review. The paper will conclude with a presentation of a case report of successful stem cell therapy for this condition.

Dilated cardiomyopathy

Cardiomyopathy is generally defined as weakening of the myocardium (Richardson *et al.*, 1996). There are three main types of cardiomyopathy: (i) dilated cardiomyopathy, which

is associated with enlargement of the left ventricle and resulting suboptimal function. This condition can be the result of hypertension, post-infarct scar tissue formation, as well as other causes such as infections, cardiotoxic drugs and in some cases pregnancy; (ii) hypertrophic cardiomyopathy is a condition in which the myocardial muscle abnormally enlarges in an asymmetric manner; this is usually attributed to genetic factors although accurate gene testing has not been developed. Hypertrophic cardiomyopathy is a known cause of sudden death in young athletes; (iii) restrictive cardiomyopathy, a condition in which the myocardium does not properly enlarge during diastole, thus not allowing for proper filling and pumping. This condition is usually caused by amyloidosis, post-transplant chronic rejection, radiation fibrosis, haemochromatosis, sarcoidosis and in some cases cardiac tumours. The subject of the current paper is dilated cardiomyopathy, which is the most prevalent of the cardiomyopathies (60%), and believed to have an annual incidence of approximately 400,000 per year in the USA alone (Karkkainen and Peuhkurinen, 2007). Although in some cases dilated cardiomyopathy occurs post-infarct as a result of ventricular wall thinning, the majority of dilated cardiomyopathies are non-ischaemia associated and believed to be caused by idiopathic, viral-induced, and parasite-induced causes. For example, in a representative study of 673 patients referred for congestive heart failure due to dilated cardiomyopathy the most common causes of dilated cardiomyopathy were idiopathic origin (47%), idiopathic myocarditis (12%), coronary artery disease (11%) and known causes such as hypertension and post-infarct remodelling (31%) (Kasper *et al.*, 1994). Although the instigating causes of cardiomyopathy are numerous and in many cases ill defined, the end result is often similar: decreased ejection fraction and progression to heart failure. Mechanisms of cardiac deterioration often include elaboration of inflammatory mediators, which cause functional impairment and apoptotic death of myocardial cells that leads to a self-feeding cascade.

Inflammatory mediators as cause of myocardial damage

Inflammation is associated with heart failure. Acute inflammation during myocardial infarction results in production of cytokines such as tumour necrosis factor (TNF) α , interleukin (IL)-1 and IL-6, which directly induce myocardial 'stunning' as well as apoptosis of contractile cells (Paulus, 2000; Caillaer *et al.*, 2004). Positive correlations have been made between the amount of TNF released into systemic circulation and the extent of myocardial cell death (Li *et al.*, 1999). In less acute circumstances, inflammatory cytokines are also known to play a role in progression to heart failure. For example, patients with non-ischaemic, as well as ischaemic dilated cardiomyopathy, possess increased concentrations of circulating TNF α compared with healthy controls (Oral *et al.*, 1999; De Biase *et al.*, 2003). Spontaneous secretion of TNF α from leukocytes in patients with dilated cardiomyopathy but not controls has also been reported (Brooksbank *et al.*, 2005). A correlative role of inflammatory cytokines in progression of heart failure is supported by studies demonstrating improved prognosis in patients with lower concentrations of TNF α and IL-6 (Hasper *et al.*, 1998; Rauchhaus *et al.*, 2000; Pan *et al.*, 2004). The possible importance of inflammatory cytokines to disease progression is seen in studies using inhibitors of inflammatory cytokines, such

as thalidomide or pentoxifyllin, which demonstrated reduction in TNF α concentrations in patients responding to treatment (Sliwa *et al.*, 2004; Orea-Tejeda *et al.*, 2006). In animal models of heart failure, blockade of TNF α (Li *et al.*, 2002; Moe *et al.*, 2004), as well as IL-1 (Thomas *et al.*, 2003) signalling, has been demonstrated to ameliorate disease progression.

While it appears that inflammation plays a key role in the progression of heart failure in general, and specifically in dilated cardiomyopathy, the next question is what causes this inflammation? In the context of infarction, it is obvious that release of intracellular components through necrosis produce a potent 'danger signal' that activates various innate immune cascades. For example, heat shock proteins (Sato *et al.*, 2006a), extracellular matrix degradation products (Samstein *et al.*, 2004), soluble mediators such as mast cell released histamine (Frangogiannis and Entman, 2006), tissue factor resulting from activation of the coagulation cascade (Frangogiannis *et al.*, 2002), and complement pathway products (Yasuda *et al.*, 1990) are all known to either directly induce inflammatory cytokine production or to elicit innate immune cell infiltration; this subsequently results in cytokine production. In more chronic conditions such as toxin-induced dilated myocarditis, it is believed that 'danger receptors' such as the toll-like receptor family can recognize tissue death and activate immune cells. For example, doxorubicin-induced cardiac dilation is associated with a chronic remodelling of the myocardium subsequent to death of a small portion of cardiomyocytes. In animals that lack toll-like receptor-2 (TLR-2), progression of disease pathology in this model is markedly inhibited in comparison to wild-type mice (Nozaki *et al.*, 2004). TLR-2 activation stimulates monocytes and dendritic cells to release inflammatory cytokines and its ligands include damage-associated proteins such as heat shock protein 60 (O'Neill, 2002; Direskeneli and Saruhan-Direskeneli, 2003). In addition, TLR-2 knockout mice also show inhibited progression to heart failure subsequent to infarct-induced chronic ventricular remodelling (Shishido *et al.*, 2003). Clinically, heart failure is associated with up-regulation of TLR-4 expression both locally in the myocardium (Frantz *et al.*, 1999) and systemically in circulating monocytes (Sato *et al.*, 2006b). Ligands of TLR-4 such as heat shock protein 70 are released by damaged myocardium and serum concentration is associated with poor prognosis (Sato *et al.*, 2006a). In addition to induction of inflammatory mediators by localized tissue damage, the simple increase of myocardial load is also associated with increased production of inflammatory cytokines by cardiomyocytes (Baumgarten *et al.*, 2002; Roncon-Albuquerque *et al.*, 2005).

Infectious causes of cardiomyopathy are described in the literature. Commonly known causes include Chagas' disease (Rodas *et al.*, 1992), coxsackie virus (Spotnitz and Lesch, 2006), and *Chlamydia pneumoniae* (Song *et al.*, 2001). Evidence exists that such diseases play an active role in stimulation of cytokine secretion, which leads to deleterious changes in the myocardium, as well as pathogens that induce initial tissue damage, which results in an autoimmune sequel. For example, Chagas' disease patients that progress to cardiomyopathy often display T-cell-mediated responses against myocardial antigens (Fuenmayor *et al.*, 2005). Systemic elevations in TNF α are also observed in patients with Chagas' disease-associated cardiomyopathy (Talvani *et al.*, 2004). The causative role of immune-mediated cardiac damage is revealed in this disease

by mice lacking IL-4 in which Chagas' infection clears at a more rapid rate as compared with wild-type mice; however cardiac pathology is exacerbated (Soares *et al.*, 2001). Similar observations linking immune attack against infectious pathogen with cardiac pathology are also seen in coxsackie virus (Fuse *et al.*, 2005) and *Chlamydia pneumoniae* models (Bachmaier and Penninger, 2005).

Numerous observations in addition to the above-mentioned studies report a strong association between ongoing inflammatory responses and progression of cardiac dysfunction. This is specifically noted in dilated cardiomyopathy where myocyte damage causes decrease in ventricular wall thickness and suppression of contractile function. In dilated cardiomyopathy, the reduction in wall thickness is associated with an elevated rate of apoptosis (Ibe *et al.*, 2007; Kietselaer *et al.*, 2007). Apoptosis is a tightly controlled process whose physiological purpose is regulating tissue mass, as well as clearing dysfunctional cells. Signals inducing programmed cell death include inflammatory cytokines, and in many cases membrane-bound molecules such as the death-inducing protein Fas ligand (FasL). In conditions of inflammation, up-regulation of myocardial Fas and FasL has been reported. For example, mice with mutations in this pathway have reduced myocardial damage in response to ischaemia (Jeremias *et al.*, 2000). A direct role of TNF α -induced Fas–FasL up-regulation has been demonstrated in myocardial cells *in vitro* (Chen and Tu, 2002). Mechanical overload (Liao *et al.*, 2005), chemical toxicity (Nitobe *et al.*, 2003; Kalivendi *et al.*, 2005), and viral infections (Seko *et al.*, 2002) have all been demonstrated to cause cardiomyocyte apoptosis through Fas–FasL either directly or indirectly through inflammatory cytokine mediated up-regulation of this pathway. Thus it appears that chronic inflammation and apoptosis form a positive feedback loop in heart failure, not only in post-infarct setting but also in chronic cardiac degeneration such as dilated cardiomyopathy.

Autologous bone marrow stem cell therapy for post-infarct and chronic ischaemia associated heart failure

The introduction of bone marrow-derived stem cells in cardiology offers hope to a patient population that previously was restricted to transplantation and palliative procedures. Additionally, bone marrow stem cells are free of ethical issues associated with embryonic stem cells (Beeson and Lippman, 2006). Initial preclinical studies have shown that bone marrow stem cells are capable of augmenting cardiac function in a post-infarct setting (Orlic *et al.*, 2001; Kudo *et al.*, 2003). Mechanistically, these cells are believed to mediate therapeutic effects through production of paracrine trophic factors (Gnecchi *et al.*, 2006; Tse *et al.*, 2007), induction of angiogenesis (Mollmann *et al.*, 2006; Shyu *et al.*, 2006), and transdifferentiation into cardiomyocytes (Orlic, 2003; Xu *et al.*, 2004; Eisenberg *et al.*, 2006). Clinical translation of bone marrow stem cell therapy has resulted in observation of reduced scar tissue, ventricular wall dilation, and increased ejection fraction after myocardial infarction (Chen *et al.*, 2004; Fernandez-Aviles *et al.*, 2004; Osterziel, 2007). Although double-blinded Phase III trials are currently in progress, the consensus is that stem cell therapy will in one form or another

play a role in the treatment of cardiac disorders. An example of stem cell trials in cardiology is presented in **Table 1**.

The attractiveness of autologous stem cell therapy comes from the fact that during cardiac infarction, stem cell mobilization to induce myocardial repair seems to be a natural response of the body. Stem cell chemoattractants such as vascular endothelial growth factor (VEGF) (Soeki *et al.*, 2000) are observed in systemic circulation subsequent to infarction. Peaking of serum VEGF at day 7 post-infarct has been correlated with mobilization of bone marrow CD34 stem cells into circulation (Shintani *et al.*, 2001). Subsequent studies have demonstrated that numerous cytokines are involved in the mobilization of endogenous bone marrow stem cells post-infarct, such as stromal-cell-derived factor-1 (SDF-1) (Deng *et al.*, 2006), granulocyte colony-stimulating factor (G-CSF) (Leone *et al.*, 2006), and IL-8 (Schomig *et al.*, 2006). Accordingly, by administering stem cells into an area proximal to cardiac infarction, one anticipates that this procedure merely serves to augment efficacy of an existing natural response. In conditions associated with chronic ischaemic heart failure, for example caused by atherosclerotic occlusions, stem cell therapy is used not only for regenerating myocardium, but also to increase circulation through formation of collateral blood vessels. Intramyocardial administration of stem cells was initially performed in 2001 as an adjuvant to bypass with angiographic evidence of collateralization at the implant site in three of five patients (Hamano *et al.*, 2001). Subsequent studies have demonstrated that bone marrow derived stem cells either as mononuclear cells (Yaoita *et al.*, 2005) or purified into subsets (Stamm *et al.*, 2007), are capable of increasing oxygenation, as well as ejection fraction when implanted into chronically ischemic myocardium. Mechanistically, in the chronic ischaemia setting, the intramuscular injection of stem cells induces various cytokine cascades that stimulate migration of endogenous endothelial precursors and interaction with the exogenously administered precursors in order to cause new blood vessel formation. It is known that chronically ischaemic tissue has activation of the transcription factor hypoxia-inducible factor 1 (HIF-1) α (Blanco Pampin *et al.*, 2006) which is associated with localized production of fibroblast growth factor-1 (FGF-1) (Iwakura *et al.*, 2000), whose role is to chemoattract the administered endothelial precursors and induce the angiogenic cascade. From the mentioned studies it is apparent that strong logic exists for the use of stem cells either as bone marrow mononuclear cells, or as purified subpopulations for repairing post-acute myocardial infarction damage, as well as chronic ischaemia. However, in chronic cardiac indications such as dilated cardiomyopathy, which represent a significant burden on our society, is there rationale for the use of stem cells?

Mesenchymal stem cell therapy for inflammatory associated heart failure

While the regenerative and angiogenic properties of stem cells are well known, another therapeutic aspect of stem cells is their anti-inflammatory properties. Bone marrow mononuclear cells are known to contain not only haematopoietic/endothelial stem cells, designated by markers such as CD34 and CD133, but also mesenchymal stem cells. For decades it was known that bone

Table 1. Sample trials of autologous stem cell cardiac clinical trials.

	<i>Institution</i>	<i>Patients treated</i>	<i>Findings</i>	<i>Reference</i>
Reperfused AMI	JW Goethe-University, Germany	59	Contrast-enhanced magnetic resonance imaging revealed an increased EF, reduced infarct size, and absence of hypertrophy	Schachinger <i>et al.</i> (2004)
Reperfused AMI	Military Medical Academy, Medicine, Belgrade	4	Significant improvement in myocardial perfusion in two patients 4 months after the infarction	Obradovic <i>et al.</i> (2004)
Reperfused AMI	Hanover Medical School, Germany	30	Increased mean global LVEF	Wollert <i>et al.</i> (2004)
Reperfused AMI	University of Frankfurt, Germany	20	Increase in global LVEF, and regional wall motion in the infarct zone, and profoundly reduced end-systolic left ventricular volumes	Assmus <i>et al.</i> (2002)
Anteroseptum AMI	Department of Cardiology, Athens	11	Improvement of myocardial contractility in one or more previously non-viable myocardial segments	Katritsis <i>et al.</i> (2005)
AMI	First Municipal Hospital, Nanjing, China	34	LVEF increased significantly compared with that preimplantation and with that of the control group at 3 months post-injection	Chen <i>et al.</i> (2004)
AMI	Liaoning Provincial People's Hospital, China	35	There was a significant improvement in global left ventricular function ejection fraction	Li ZQ <i>et al.</i> (2007)
AMI (recent and old)	Seoul National University	41	Significant improvement in LVEF and remodelling compared with controls	Kang <i>et al.</i> (2006)
Ischaemic cardiomyopathy	University of Pittsburgh	10	The ejection fractions of the untreated versus treated: pre-operative	Patel <i>et al.</i> (2005)
Ischaemic cardiomyopathy	Ege University, Turkey	6	There was a significant increase in life quality and NYHA class; some benefit was documented on echocardiography, scintigraphy and PET	Ozbaran <i>et al.</i> (2004)
End-stage heart failure	Navy General Hospital, Beijing	14	LVEF increased; left ventricular end-systolic volume decreased	Gao <i>et al.</i> (2006)
Refractory angina	Leiden University Medical Center	25	CCS and Quality of life improved; LVEF increased	Beeres <i>et al.</i> (2006)
Refractory angina	San Raffaele Hospital, Milan, Italy	10	Severity of angina improved by ≥ 2 classes in three patients and quality of life in all patients; myocardial perfusion improved in four of eight patients	Briguori <i>et al.</i> (2006)
Chronic myocardial ischaemia	Washington Hospital Center	10	CCS improved, as well as stress-induced ischaemia occurring within the injected territories	Fuchs <i>et al.</i> (2003)
Stable ischaemic heart disease	Goethe University, Frankfurt	75	Significant improvement in the left ventricular ejection fraction after 3 months	Assmus <i>et al.</i> (2006)
Dilated cardiomyopathy	Centro Medico Nacional Siglo XXI, Mexico	39	Pre-operatively, the functional class for 26 of these patients was III; post-operatively, functional classes were II in five cases and I in 15 patients	Arguero <i>et al.</i> (2006)

AMI = acute myocardial infarction; CCS = Cleveland Clinic Score; EF = ejection fraction; LVEF = left ventricular ejection fraction; NYHA = New York Heart Association; PET = positron emission tomography.

marrow cells contain immune regulatory cells. For example, in the 1970s, bone marrow-derived 'natural suppressor' cells were demonstrated to possess antigen-non-specific immune regulatory activity (Adler *et al.*, 1976). These cells were subsequently found to secrete large amounts of transforming growth factor (TGF) β , which inhibits T cell and antigen-presenting cell activity (Moore *et al.*, 1992). Subsequent studies have found that mesenchymal stem cells of the bone marrow compartment also produce large amounts of this cytokine and can mediate potent immune suppressive activity *in vitro* and *in vivo* (Li H *et al.* 2007).

Currently, expanded bone marrow derived allogeneic mesenchymal stem cells are being used in Phase III clinical trials for treatment of the inflammatory conditions graft versus host disease and Crohn disease (<http://www.osiris.com>, accessed 18 January 2008). A recently completed Phase I trial demonstrated profound activity of these cells in preserving left ventricular ejection fraction in patients following myocardial infarction (http://files.shareholder.com/downloads/OSIR/280683646x0x86980/8a03c252-36c7-412d-8c9f-0526e0de0993/OSIR_News_2007_3_25_General.pdf). The authors believe that in conditions such as dilated cardiomyopathy, which possess a chronic inflammatory component, the use of mesenchymal stem cells may be more optimal than bone marrow stem cells alone. This rationale is based on findings that while mesenchymal stem cells exert similar functions as bone marrow stem cells such as induction of cardiac angiogenesis (Tang *et al.*, 2006), prevention of myocardial apoptosis (Gnecchi *et al.*, 2006), and inhibition pathological cardiac remodelling associated with left ventricle dilation and wall thinning (Berry *et al.*, 2006), these cells also have a potent anti-inflammatory component. Specifically, mesenchymal stem cells have been shown to inhibit a wide variety of chronic inflammatory conditions such as experimental allergic encephalomyelitis (Zappia *et al.*, 2005), collagen-induced arthritis (Djouad *et al.*, 2005), and protect from immune-mediated diabetes (Lee *et al.*, 2006). Mesenchymal stem cells have also been shown to inhibit myocardial specific inflammation including TNF α production, matrix-metalloproteinase activation, and release of scar tissue formation (Guo *et al.*, 2007). Furthermore, mesenchymal stem cells have been demonstrated effective in animal models of dilated cardiomyopathy (Nagaya *et al.*, 2005). An added advantage of mesenchymal stem cell therapy is that these cells have demonstrated efficacy by intravenous administration both in animal models of cardiac failure (Boomsma *et al.*, 2006), as well as in clinical trials (http://files.shareholder.com/downloads/OSIR/280683646x0x86980/8a03c252-36c7-412d-8c9f-0526e0de0993/OSIR_News_2007_3_25_General.pdf).

Another reason supporting the use of mesenchymal stem cells as opposed to bone marrow stem cells alone are findings that stem cells from older patients have reduced homing activity *in vitro* as compared with stem cells from younger patients. Importantly, stem cells from patients with heart disease exhibit profoundly reduced neovascularization and angiogenic ability when administered to immune deficient mice with experimental ischemia (Heeschen *et al.*, 2004). The use of allogeneic mesenchymal stem cells allows for

selection of 'younger' donor sources without the limitation of the autologous donor age.

Case report: placental matrix-derived mesenchymal stem cells and cord blood CD34 cells in treatment of dilated cardiomyopathy

The use of cord blood stem cells in the absence of recipient conditioning has previously been reported by Ghen *et al.* (2006) for the treatment of amyotrophic lateral sclerosis with some degree of success. Additionally, numerous other reports have been published in which cord blood and/or placentally derived cells have been administered across allogeneic barriers without adverse effect, reviewed in Ghen *et al.*, (2006). In order to augment the number of potentially therapeutic stem cells from this source, the authors' laboratory has developed methods of expanding mesenchymal stem cells from placental matrix. This source of stem cells is potentially angiogenic and possesses higher regenerative activity as compared with mesenchymal stem cells from bone marrow and adipose tissue (Kern *et al.*, 2006; Musina *et al.*, 2006; Edwards and Hollands, 2007). Additionally, several technologies have been developed for expansion of cord blood-derived CD34 stem cells using a serum-free protocol. All placental material used for these investigations was collected in a sterile manner with informed consent of the donor. Here, a case of a 50-year-old patient with dilated cardiomyopathy treated with this combination is reported.

The patient was diagnosed with dilated, non-ischaemic cardiomyopathy in February 2002, with an ejection fraction of 30% as measured by echocardiogram (ECHO). The clinical presentation at diagnosis was indicative of congestive heart failure, including marked dyspnoea, inability to exercise, dizziness, and irregular heart beat; New York Heart Association (NYHA) classification of II (see http://www.abouthf.org/questions_stages.htm). ECHO analysis in April 2003 indicated ejection fraction of approximately 40%. Consultation in December of 2005 revealed ejection fraction remained at 30–40% and the patient was placed on carvedilol 25 mg p.o. b.i.d., valsartan 325 mg p.o. q.d., amlodipine besilate 10 mg p.o. q.d., hydrochlorothiazide 25 mg/day, aldosterone, and acetylsalicylic acid 81 mg/day. Weekly blood pressure measurements from the middle of November to the middle of December 2005 were: 145/100, 150/100, 145/90 and 150/95. Quality of life assessment using the Minnesota Living with Heart Failure Questionnaire (Middel *et al.*, 2001) revealed a score of 90. The Minnesota Living with Heart Failure Questionnaire consists of 21 questions to assess a patient's perception of how he or she is affected by heart failure, physically and emotionally. The patient presented in December 2006 and subsequent to being explained the experimental nature of the therapy proposed, the patient signed informed consent. At the time of the treatment the patient had a NYHA classification of III and was taking the above medications, low-dose human growth hormone, testosterone, and the following dietary supplements: co-enzyme Q10, magnesium, L-carnitine, omega 3, 6 and 9 fatty acids, and D-ribose. The patient was treated with cord blood expanded allogeneic CD34 cells (2.5 million) and placentally derived allogeneic mesenchymal stem cells (3 million) three times over the period of a week. Cellular therapy was well tolerated

and no adverse side effects were observed either acutely or as of this writing, 11 months post-treatment. Notably the patient did not experience either symptoms of either acute (skin rash of diarrhoea) or chronic (skin rash, skin inflammation, mouth lesions, hair loss, indigestion) graft- versus host- disease. Two weeks prior to an ECHO in April 2007, the patient voluntarily discontinued all above-mentioned medications and supplements. The ECHO revealed an ejection fraction of 50–55%. The patient reports profound clinical benefit at time of writing (November 2007), including resolution of heart-failure-associated symptoms of dizziness, fatigue, dyspnea, rapid heart beat, irregular heart beat, depression, blackouts, and loss of sleep secondary to orthopnoea. The Minnesota Living with Heart Failure Questionnaire score was zero. The patient has a normal ejection fraction and no symptoms of heart failure and is no longer classifiable on the NYHA scale. Weekly blood pressure measurements for the month of April 2007 were 120/80, 110/85, 120/90 and 110/86. Given that the patient was treated under compassionate-use exemptions and not as part of a clinical trial, no blood gas or pulmonary function data were collected. Although no firm conclusions can be drawn from a single patient case report, and no cytokine analysis was performed to demonstrate potential anti-inflammatory effects of stem cells, these findings suggest that stem cell therapy using CD34 and mesenchymal cells should be investigated in larger groups of cardiomyopathy patients under controlled settings.

Declaration

Neil Riordan and Thomas Ichim own stocks in Medistem Inc. (mdsm.ob) and are involved in the company's day-to-day operations.

References

- Adler S, Singhal SK, Sercarz EE. 1976 Regulatory cells in the bone marrow. *Advances in Experimental Medicine and Biology* **66**, 599–605.
- Arguero R, Careaga-Reyna G, Castaño-Guerra R et al. 2006 Cellular autotransplantation for ischemic and idiopathic dilated cardiomyopathy. Preliminary report. *Archives of Medical Research* **37**, 1010–1014.
- Assmus B, Honold J, Schächinger V et al. 2006 Transcoronary transplantation of progenitor cells after myocardial infarction. *New England Journal of Medicine* **355**, 1222–1232.
- Assmus B, Schächinger V, Teupe C et al. 2002 Transplantation of progenitor cells and regeneration enhancement in acute myocardial infarction (TOPCARE-AMI). *Circulation* **106**, 3009–3017.
- Bachmaier, K, Penninger JM 2005 Chlamydia and antigenic mimicry. *Current Topics in Microbiology and Immunology* **296**, 153–163.
- Baumgarten G, Knuefermann P, Kalra D et al. 2002 Load-dependent and -independent regulation of proinflammatory cytokine and cytokine receptor gene expression in the adult mammalian heart. *Circulation* **105**, 2192–2197.
- Beeres SL, Bax JJ, Dibbets-Schneider P et al. 2006 Sustained effect of autologous bone marrow mononuclear cell injection in patients with refractory angina pectoris and chronic myocardial ischemia: twelve-month follow-up results. *American Heart Journal* **152**, 684 e11–16.
- Beeson, D, Lippman A 2006 Egg harvesting for stem cell research: medical risks and ethical problems. *Reproductive BioMedicine Online* **13**, 573–579.
- Berry MF, Engler AJ, Woo WJ et al. 2006 Mesenchymal stem cell injection after myocardial infarction improves myocardial compliance. *American Journal of Physiology Heart and Circulatory Physiology*. **290**, H2196–H2203.
- Blanco Pampin J, Garcia Rivero SA, Otero Cepeda XL et al. 2006 Immunohistochemical expression of HIF-1alpha in response to early myocardial ischemia. *Journal of Forensic Sciences* **51**, 120–124.
- Boomsma RA., Swaminathan PD, Geenen DL et al. 2006 Intravenously injected mesenchymal stem cells home to viable myocardium after coronary occlusion and preserve systolic function without altering infarct size. *International Journal of Cardiology* **122**, 17–28.
- Briguori C, Reimers B, Sarais C et al. 2006 Direct intramyocardial percutaneous delivery of autologous bone marrow in patients with refractory myocardial angina. *American Heart Journal* **151**, 674–680.
- Brooksbank R, Woodiwiss A, Sliwa K et al. 2005 Sustained white cell cytokine activation in idiopathic dilated cardiomyopathy despite haemodynamic improvement with medical therapy. *Cardiovascular Journal of South Africa* **16**, 200–204.
- Cailleret M, Amadou A, Andrieu-Abadie N et al. 2004 N-Acetylcysteine prevents the deleterious effect of tumor necrosis factor-(alpha) on calcium transients and contraction in adult rat cardiomyocytes. *Circulation* **109**, 406–411.
- Chen QM, Tu VC 2002 Apoptosis and heart failure: mechanisms and therapeutic implications. *American Journal of Cardiovascular Drugs* **2**, 43–57.
- Chen SL, Fang WW, Qian J et al. 2004 Improvement of cardiac function after transplantation of autologous bone marrow mesenchymal stem cells in patients with acute myocardial infarction. *Chinese Medical Journal* **117**, 1443–1448.
- De Biase L, Pignatelli P, Lenti L et al. 2003 Enhanced TNF alpha and oxidative stress in patients with heart failure: effect of TNF alpha on platelet O₂⁻ production. *Journal of Thrombosis and Haemostasis* **90**, 317–325.
- Deng ZR, Yang C, Ma AQ et al. 2006 [Dynamic changes of plasma VEGF, SDF-1 and peripheral CD34+ cells in patients with acute myocardial infarction]. *Nan Fang Yi Ke Da Xue Xue Bao* **26**, 1637–1640.
- Direskeneli H, Saruhan-Direskeneli G 2003 The role of heat shock proteins in Behçet's disease. *Clinical and Experimental Rheumatology* **21**, S44–48.
- Djouad F, Fritz V, Apparailly F et al. 2005 Reversal of the immunosuppressive properties of mesenchymal stem cells by tumor necrosis factor alpha in collagen-induced arthritis. *Arthritis and Rheumatism* **52**, 1595–1603.
- Edwards RG, Hollands P 2007 Will stem cells in cord blood, amniotic fluid, bone marrow and peripheral blood soon be unnecessary in transplantation? *Reproductive BioMedicine Online* **14**, 396–401.
- Eisenberg CA, Burch JB, Eisenberg LM 2006 Bone marrow cells transdifferentiate to cardiomyocytes when introduced into the embryonic heart. *Stem Cells* **24**, 1236–1245.
- Fernandez-Aviles F, San Roman JA, García-Frade J et al. 2004 Experimental and clinical regenerative capability of human bone marrow cells after myocardial infarction. *Circulation Research* **95**, 742–748.
- Findikli N, Candan NZ, Kahraman S 2006 Human embryonic stem cell culture: current limitations and novel strategies. *Reproductive BioMedicine Online* **13**, 581–590.
- Frangogiannis NG, Entman ML 2006 Identification of mast cells in the cellular response to myocardial infarction. *Methods in Molecular Biology* **315**, 91–101.
- Frangogiannis NG, Smith CW, Entman ML 2002 The inflammatory response in myocardial infarction. *Cardiovascular Research* **53**, 31–47.
- Frantz, S, Kobzik L, Kim YD et al. 1999 Toll4 (TLR4) expression in cardiac myocytes in normal and failing myocardium. *Journal of Clinical Investigation* **104**, 271–280.
- Fuchs S, Satler LF, Kornowski R et al. 2003 Catheter-based autologous bone marrow myocardial injection in no-option patients with advanced coronary artery disease: a feasibility study. *Journal of the American College of Cardiologists* **41**, 1721–1724.
- Fuenmayor C, Higuchi ML, Carrasco H et al. 2005 Acute Chagas' disease: immunohistochemical characteristics of T cell infiltrate

- and its relationship with *T. cruzi* parasitic antigens. *Acta Cardiologica* **60**, 33–37.
- Fuse K, Chan G, Liu Y et al. 2005 Myeloid differentiation factor-88 plays a crucial role in the pathogenesis of coxsackievirus B3-induced myocarditis and influences type I interferon production. *Circulation* **112**, 2276–2285.
- Gao LR, Wang ZG, Zhu ZM et al. 2006 Effect of intracoronary transplantation of autologous bone marrow-derived mononuclear cells on outcomes of patients with refractory chronic heart failure secondary to ischemic cardiomyopathy. *American Journal of Cardiology* **98**, 597–602.
- Ghen MJ, Roshan R, Roshan RO et al. 2006 Potential clinical applications using stem cells derived from human umbilical cord blood. *Reproductive BioMedicine Online* **13**, 562–572.
- Gnecchi M, He H, Noiseux N et al. 2006 Evidence supporting paracrine hypothesis for Akt-modified mesenchymal stem cell-mediated cardiac protection and functional improvement. *FASEB Journal* **20**, 661–669.
- Guo J, Lin GS, Bao CY et al. 2007 Anti-inflammation role for mesenchymal stem cells transplantation in myocardial infarction. *Inflammation* **30**, 97–104.
- Hamano K, Nishida M, Hirata K et al. 2001 Local implantation of autologous bone marrow cells for therapeutic angiogenesis in patients with ischemic heart disease: clinical trial and preliminary results. *Japanese Circulation Journal* **65**, 845–847.
- Hasper D, Hummel M, Kleber FX et al. 1998 Systemic inflammation in patients with heart failure. *European Heart Journal* **19**, 761–765.
- Heeschen C, Lehmann R, Honold J et al. 2004 Profoundly reduced neovascularization capacity of bone marrow mononuclear cells derived from patients with chronic ischemic heart disease. *Circulation* **109**, 1615–1622.
- Ibe W, Saraste A, Lindemann S et al. 2007 Cardiomyocyte apoptosis is related to left ventricular dysfunction and remodeling in dilated cardiomyopathy, but is not affected by growth hormone treatment. *European Journal of Heart Failure* **9**, 160–167.
- Iwakura A, Fujita M, Ikemoto M et al. 2000 Myocardial ischemia enhances the expression of acidic fibroblast growth factor in human pericardial fluid. *Heart Vessels* **15**, 112–116.
- Jeremias I, Kupatt C, Martin-Villalba A et al. 2000 Involvement of CD95/Apo1/Fas in cell death after myocardial ischemia. *Circulation* **102**, 915–920.
- Kalivendi SV, Konorev EA, Cunningham S et al. 2005 Doxorubicin activates nuclear factor of activated T-lymphocytes and Fas ligand transcription: role of mitochondrial reactive oxygen species and calcium. *Biochemistry Journal* **389**, 527–539.
- Kang HJ, Lee HY, Na SH et al. 2006 Differential effect of intracoronary infusion of mobilized peripheral blood stem cells by granulocyte colony-stimulating factor on left ventricular function and remodeling in patients with acute myocardial infarction versus old myocardial infarction: the MAGIC Cell-3-DES randomized, controlled trial. *Circulation* **114** (Suppl. 1), I145–I151.
- Karkkainen S, Peuhkurinen K 2007 Genetics of dilated cardiomyopathy. *Annals of Medicine* **39**, 91–107.
- Kasper EK, Agema WR, Hutchins GM et al. 1994 The causes of dilated cardiomyopathy: a clinicopathologic review of 673 consecutive patients. *Journal of the American College of Cardiology* **23**, 586–590.
- Katristsis DG, Sotiropoulou PA, Karvouni E et al. 2005 Transcatheter transplantation of autologous mesenchymal stem cells and endothelial progenitors into infarcted human myocardium. *Catheterization and Cardiovascular Interventions* **65**, 321–329.
- Kern S, Eichler H, Stoeve J et al. 2006 Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue. *Stem Cells* **24**, 1294–1301.
- Kietselaer BL, Reutelingsperger CP, Boersma HH et al. 2007 Noninvasive detection of programmed cell loss with 99mTc-labelled annexin A5 in heart failure. *Journal of Nuclear Medicine* **48**, 562–567.
- Kudo M, Wang Y, Wami MA et al. 2003 Implantation of bone marrow stem cells reduces the infarction and fibrosis in ischemic mouse heart. *Journal of Molecular and Cellular Cardiology* **35**, 1113–1119.
- Lee RH, Seo MJ, Reger RL et al. 2006 Multipotent stromal cells from human marrow home to and promote repair of pancreatic islets and renal glomeruli in diabetic NOD/SCID mice. *Proceedings of the National Academy of Sciences of the USA* **103**, 17438–17443.
- Leone AM, Rutella S, Bonanno G et al. 2006 Endogenous G-CSF and CD34+ cell mobilization after acute myocardial infarction. *International Journal of Cardiology* **111**, 202–208.
- Li D, Zhao L, Liu M et al. 1999 Kinetics of tumor necrosis factor alpha in plasma and the cardioprotective effect of a monoclonal antibody to tumor necrosis factor alpha in acute myocardial infarction. *American Heart Journal* **137**, 1145–1152.
- Li H, Guo ZK, Li XS et al. 2007 Functional and phenotypic alteration of intrasplenic lymphocytes affected by mesenchymal stem cells in a murine allosplenocyte transfusion model. *Cell Transplantation* **16**, 85–95.
- Li W, Gan R, Sun G 2002 Chronic treatment of enbrel in rats with isoproterenol-induced congestive heart failure limits left ventricular dysfunction and remodeling. *Chinese Medical Journal* **115**, 1166–1169.
- Li ZQ, Zhang M, Jing YZ et al. 2007 The clinical study of autologous peripheral blood stem cell transplantation by intracoronary infusion in patients with acute myocardial infarction (AMI). *International Journal of Cardiology* **115**, 52–56.
- Liao X, Wang X, Gu Y et al. 2005 Involvement of death receptor signaling in mechanical stretch-induced cardiomyocyte apoptosis. *Life Sciences* **77**, 160–174.
- Middel B, Bouma J, de Jongste M et al. 2001 Psychometric properties of the Minnesota Living with Heart Failure Questionnaire (MLHF-Q). *Clinical Rehabilitation* **15**, 489–500.
- Moe GW, Marin-Garcia J, Konig A et al. 2004 In vivo TNF-alpha inhibition ameliorates cardiac mitochondrial dysfunction, oxidative stress, and apoptosis in experimental heart failure. *American Journal of Physiology Heart and Circulatory Physiology* **287**, H1813–20.
- Mollmann H, Nef HM, Kosin S et al. 2006 Bone marrow-derived cells contribute to infarct remodelling. *Cardiovascular Research* **71**, 661–671.
- Moore SC, Shaw MA, Soderberg LS. 1992 Transforming growth factor-beta is the major mediator of natural suppressor cells derived from normal bone marrow. *Journal of Leukocyte Biology* **52**, 596–601.
- Musina RA, Bekchanova ES, Belyavskii AV, Sukhikh GT 2006 Differentiation potential of mesenchymal stem cells of different origin. *Bulletin of Experimental Biology and Medicine* **141**, 147–151.
- Nagaya N, Kangawa K, Itoh T et al. 2005 Transplantation of mesenchymal stem cells improves cardiac function in a rat model of dilated cardiomyopathy. *Circulation* **112**, 1128–1135.
- Nitobe J, Yamaguchi S, Okuyama M et al. 2003 Reactive oxygen species regulate FLICE inhibitory protein (FLIP) and susceptibility to Fas-mediated apoptosis in cardiac myocytes. *Cardiovascular Research* **57**, 119–128.
- Nozaki N, Shishido T, Takeishi, et al. 2004 Modulation of doxorubicin-induced cardiac dysfunction in toll-like receptor-2-knockout mice. *Circulation* **110**, 2869–2874.
- O'Neill LA 2002 Toll-like receptor signal transduction and the tailoring of innate immunity: a role for Mal? *Trends in Immunology* **23**, 296–300.
- Obradovic S, Rusovic S, Balint B et al. 2004 Autologous bone marrow-derived progenitor cell transplantation for myocardial regeneration after acute infarction. *Vojnosanit Pregl* **61**, 519–529.
- Oral H, Fisher SG, Fay WP et al. 1999 Effects of amiodarone on tumor necrosis factor-alpha levels in congestive heart failure secondary to ischemic or idiopathic dilated cardiomyopathy. *American Journal of Cardiology* **83**, 388–391.
- Orea-Tejada A, Arrieta-Rodriguez O, Castillo-Martinez L et al. 2006 Effects of thalidomide treatment in heart failure patients. *Cardiology* **108**, 237–242.
- Orlic D 2003 Adult bone marrow stem cells regenerate myocardium

- in ischemic heart disease. *Annals of the New York Academy of Sciences USA* **996**, 152–157.
- Orlic D, Kajstura J, Chimenti S et al. 2001 Bone marrow cells regenerate infarcted myocardium. *Nature* **410**, 701–705.
- Osterziel KJ 2007 Improved clinical outcome after intracoronary administration of bone-marrow-derived progenitor cells in acute myocardial infarction: final 1 year results of the REPAIR-AMI trial. *European Heart Journal* **28**, 638.
- Ozbaran M, Omay SB, Nalbantgil S et al. 2004 Autologous peripheral stem cell transplantation in patients with congestive heart failure due to ischemic heart disease. *European Journal of Cardiothoracic Surgery* **25**, 342–50; discussion 350–1.
- Pan JP, Liu TY, Chiang SC et al. 2004 The value of plasma levels of tumor necrosis factor-alpha and interleukin-6 in predicting the severity and prognosis in patients with congestive heart failure. *Journal of the Chinese Medical Association* **67**, 222–228.
- Patel AN, Geffner L, Vina RF et al. 2005 Surgical treatment for congestive heart failure with autologous adult stem cell transplantation: a prospective randomized study. *Journal of Thoracic and Cardiovascular Surgery* **130**, 1631–1638.
- Patel WJ 2000 Cytokines and heart failure. *Heart Failure Monitor* **1**, 50–56.
- Rauchhaus M, Koloczek V, Volk H et al. 2000 Inflammatory cytokines and the possible immunological role for lipoproteins in chronic heart failure. *International Journal of Cardiology* **76**, 125–133.
- Richardson P, McKenna W, Bristow M et al. 1996 Report of the 1995 World Health Organization/International Society and Federation of Cardiology Task Force on the Definition and Classification of cardiomyopathies. *Circulation* **93**, 841–842.
- Rodas A, Toro S, Ramos A et al. 1992 [The incidence of *Trypanosoma cruzi* antibodies in patients with dilated cardiomyopathy at the Instituto Nacional de Cardiologia Ignacio Chavez]. *Archives of the Institute of Cardiology of Mexico* **62**, 541–545.
- Roncon-Albuquerque RJ, Vasconcelos M, Lourenço AP et al. 2005 Activation profile of pro-inflammatory cytokines in acute cardiac overload. *Revista Portuguesa de Cardiologia* **24**, 1369–1378.
- Samstein B, Johnson GB, Platt JL 2004 Toll-like receptor-4 and allograft responses. *Transplantation* **77**, 475–477.
- Satoh M, Shimoda Y, Akatsu T et al. 2006a Elevated circulating levels of heat shock protein 70 are related to systemic inflammatory reaction through monocyte Toll signal in patients with heart failure after acute myocardial infarction. *European Journal of Heart Failure* **8**, 810–815.
- Satoh M, Shimoda Y, Maesawa C et al. 2006b Activated toll-like receptor 4 in monocytes is associated with heart failure after acute myocardial infarction. *International Journal of Cardiology* **109**, 226–234.
- Schachinger V, Assmus Britten MB et al. 2004 Transplantation of progenitor cells and regeneration enhancement in acute myocardial infarction: final one-year results of the TOPCARE-AMI Trial. *Journal of the American College of Cardiology* **44**, 1690–1699.
- Schomig K, Busch G, Steppich B et al. 2006 Interleukin-8 is associated with circulating CD133+ progenitor cells in acute myocardial infarction. *European Heart Journal* **27**, 1032–1037.
- Seko Y, Kayagaki N et al. 2002 Role of Fas/FasL pathway in the activation of infiltrating cells in murine acute myocarditis caused by Coxsackievirus B3. *Journal of the American College of Cardiology* **39**, 1399–1403.
- Shintani S, Murohara T, Ikeda H et al. 2001 Mobilization of endothelial progenitor cells in patients with acute myocardial infarction. *Circulation* **103**, 2776–2779.
- Shishido T, Nozaki N, Yamaguchi S et al. 2003 Toll-like receptor-2 modulates ventricular remodeling after myocardial infarction. *Circulation* **108**, 2905–2910.
- Shyu KG, Wang BW, Hung HF et al. 2006 Mesenchymal stem cells are superior to angiogenic growth factor genes for improving myocardial performance in the mouse model of acute myocardial infarction. *Journal of Biomedical Sciences* **13**, 47–58.
- Sliwa K, Woodiwiss A, Kone VN et al. 2004 Therapy of ischemic cardiomyopathy with the immunomodulating agent pentoxifylline: results of a randomized study. *Circulation* **109**, 750–755.
- Soares MB, Silva-Mota KN, Lima RS et al. 2001 Modulation of chagasic cardiomyopathy by interleukin-4: dissociation between inflammation and tissue parasitism. *American Journal of Pathology* **159**, 703–709.
- Soeki T, Tamura Y, Shinohara H et al. 2000 Role of circulating vascular endothelial growth factor and hepatocyte growth factor in patients with coronary artery disease. *Heart Vessels* **15**, 105–111.
- Song H, Takaki H, Yashiro A et al. 2001 Dilated cardiomyopathy and *Chlamydia pneumoniae* infection. *Heart* **86**, 456–458.
- Spotnitz MD, Lesch M 2006 Idiopathic dilated cardiomyopathy as a late complication of healed viral (coxsackie B virus) myocarditis: historical analysis, review of the literature, and a postulated unifying hypothesis. *Progress in Cardiovascular Diseases* **49**, 42–57.
- Stamm C, Kleine HD, Choi YH et al. 2007 Intramyocardial delivery of CD133+ bone marrow cells and coronary artery bypass grafting for chronic ischemic heart disease: safety and efficacy studies. *Journal of Thoracic and Cardiovascular Surgery* **133**, 717–725.
- Talvani A, Rocha MO, Barcelos LS et al. 2004 Elevated concentrations of CCL2 and tumor necrosis factor-alpha in chagasic cardiomyopathy. *Clinical Infectious Diseases* **38**, 943–950.
- Tang J, Xie Q, Pan G et al. 2006 Mesenchymal stem cells participate in angiogenesis and improve heart function in rat model of myocardial ischemia with reperfusion. *European Journal of Cardiothoracic Surgery* **30**, 353–361.
- Thomas JA, Haudek SB, Koroglu T et al. 2003 IRAK1 deletion disrupts cardiac Toll/IL-1 signaling and protects against contractile dysfunction. *American Journal of Physiology Heart and Circulatory Physiology* **285**, H597–H606.
- Tse HF, Siu CW, Zhu SG et al. 2007 Paracrine effects of direct intramyocardial implantation of bone marrow derived cells to enhance neovascularization in chronic ischaemic myocardium. *European Journal of Heart Failure* **9**, 747–753.
- van de Stolpe A, van den Brink S, van Rooijen M et al. 2005 Human embryonic stem cells: towards therapies for cardiac disease. Derivation of a Dutch human embryonic stem cell line. *Reproductive BioMedicine Online* **11**, 476–485.
- Wollert KC, Meyer GP, Lotz J et al. 2004 Intracoronary autologous bone-marrow cell transfer after myocardial infarction: the BOOST randomised controlled clinical trial. *Lancet* **364**(9429), 141–148.
- Xu M, Wani M, Dai YS et al. 2004 Differentiation of bone marrow stromal cells into the cardiac phenotype requires intercellular communication with myocytes. *Circulation* **110**, 2658–2665.
- Yaoita H, Takase S, Maruyama Y et al. 2005 Scintigraphic assessment of the effects of bone marrow-derived mononuclear cell transplantation combined with off-pump coronary artery bypass surgery in patients with ischemic heart disease. *Journal of Nuclear Medicine* **46**, 1610–1617.
- Yasuda M, Takeuchi K, Hiruma et al. 1990 The complement system in ischemic heart disease. *Circulation* **81**, 156–163.
- Zappia E, Casazza S, Pedemonte E et al. 2005 Mesenchymal stem cells ameliorate experimental autoimmune encephalomyelitis inducing T-cell anergy. *Blood* **106**, 1755–1761.

Received 8 August 2007; refereed 27 September 2007; accepted 14 January 2008.